JCU ePrints

This file is part of the following reference:

Shinzato, Chuya (2007) Cnidarian Sox genes and the evolution of function in the Sox gene family. PhD thesis, James Cook University.

Access to this file is available from:

http://eprints.jcu.edu.au/7948



Cnidarian Sox genes and the evolution of function in the Sox gene family

Thesis Submitted by

Chuya SHINZATO, Master of Agriculture, Kyoto University, Japan

in October 2007

Thesis submitted in fulfillment of the requirements of the degree of Doctor of Philosophy in the School of Pharmacy and Molecular Sciences at James Cook University

Statement of Sources		
I declare that this thesis is my own work and hanother degree or diploma at any university or Information derived from the published or wacknowledged in the text and a list of references	other institution of tertian other institution of other of other o	ry education.
Chuya SHINZATO		
Signature	Date	

Acknowledgements

First of all, I am sorry for my poor English. Because English is not my first language, it is very difficult for me to express my appreciation, no matter how much I appreciate everybody.

Firstly, I would also like to thank to my supervisor Prof. David Miller for giving me the opportunity to do PhD. Without his encouragement, I could not get PhD. I also thank Dr. Eldon Ball for fruitful discussion and suggestions. I thank Dr. David Hayward for his advices and sending me samples. I thank all past and present members of the DM lab.

I thank people who have supported (financially and mentally) my PhD studying in Australia, especially my family and friends.

I also wish to acknowledge two years financial support from the Okinawa International Exchange & Human Resources Development Foundation.

Finally, I dedicate this thesis to my grand father, Shinsyo Arakaki. Without his one-year financial help, I had to give up continuing my PhD study here in Townsville.

Table of Contents

Statement of Access	i
Statement of Sources	ii
Acknowledgements	iii
Abstract	xi
List of Figures	vii
List of Tables	X
Chapter 1 – General Introduction	1
1.1 Importance of the coral <i>Acropora millepora</i> as a simple and basal metazoan	1
1.1.1 Unexpected gene complexity and conserved functions in cnidarians	2
1.2 Sox genes	5
1.3 Cnidarian Bilogy	13
1.3.1 Early embryogenesis in cnidarians, focusing on A. millepora	13
1.3.2 Cnidarian sex determination	19
1.3.3 Cnidarian nervous system	20
1.4 Project objectives	21
Chapter 2 – The Sox gene family in the <i>Acropora millepora</i>	23
2.1 Introduction	23
2.2 Materials and Methods	23
2.2.1 Animal sampling	23
2.2.2 Isolation of Sox genes from Acropora millepora	23
2.2.3 Phylogenetic analysis	24
2.2.4 Fixation and whole mount in situ hybridization of coral embryo	25
2.2.5 Whole mount in situ hybridization for Nematostella vectensis	26
2.3 Results	27
2.3.1 The Acropora Sox gene complement	27
2.3.2 The Acropora SoxB and SoxE genes are expressed in the presumptive	
ectoderm and endoderm respectively during gastrulation	32
2.3.3 AmSoxC is expressed in sensory neurons	35
2.3.4 Expression analysis of AmSoxB2 and AmSoxF	41
2.4 Discussion	42

2.4.1 Comparison of the Acropora and Nematostella Sox complements	42
2.4.2 Anthozoan Sox genes and the evolutionary divergence of the Sox family	42
2.4.3 Expression patterns of <i>Acropora</i> Sox genes – glimpses of ancestral	
functions?	43
2.4.4 Heterogeneity in early Sox expression patterns within the Anthozoa	45
Chapter 3 -A molecular mechanism for early embryogenesis in Acropora millepora	: are
Sox genes involved in the canonical Wnt/β-catenin pathway?	48
3.1 Introduction	48
3.2 Materials and methods	54
3.2.1 Alsterpaullone treatment	54
3.2.2 Whole mount immunohistochemistry	54
3.2.3 Western blots	55
3.2.4 RT-PCR	55
3.3 Results	57
3.3.1 Spatial expression pattern of Ambcat mRNA during early embryogenesis	57
3.3.2 Alsterpaullone inhibits <i>Acropora</i> gastrulation	59
3.3.3 Distribution of <i>Acropora</i> β-catenin protein during early embryogenesis	61
3.3.4 AmSoxB1 is suppressed by alsterpaullone treatment	66
3.3.5 Zygotic expression of AmSoxE1 is suppressed by alsterpaullone treatment	nt 69
3.3.6 Alsterpaullone does not affect AmSoxB2, AmSoxB3, AmSoxC, or hbn-A	\ m
expression	71
3.3.7 Alsterpaullone induces widespread <i>Ambcat</i> expression	71
3.4 Discussion	74
3.4.1 The distribution of <i>Acropora</i> β-catenin protein during early embryogenes	sis
is not conserved in other animals	74
3.4.2 AmSoxB1 and AmSoxE1 are potential downstream targets of β-catenin	
signaling	77
Chapter 4 - General Conclusions	80
4.1 Highly conserved <i>Acropora</i> Sox genes	80
4.2 Does <i>Acropora</i> provide insights into the ancestral functions of Sox genes?	81

4.3 No simple relationship between molecular developmental r	nechanisms of
cnidarians and those of bilaterians	82
4.4 Future directions	85
References	87
Appendix A – Abbreviations	97
Appendix B – Supplementary data	99
Supplementary figures	100
Supplementary method	117
Supplementary table	117

List of Figures

Figure 1.1	Evolutionary relationships of the Metazoa and Cnidaria.	2
Figure 1.2	Model Anthozoans	5
-	Schematic representation of SOX proteins highlighting conservation within y groups	
	A conserved regulation of neural specific Sox genes by BMP-antagonist	9
Figure 1.5	A conserved transcriptional pathway in endoderm differentiation	0
Figure 1.6	Huge diversity of cnidarian gastrulation mechanisms	4
	Schematic representations of the formation of the two germ layers during n in <i>Acropora</i>	5
Figure 1.8	Micrographs of the embryonic development of Acropora millepora	8
	Maximum Likelihood phylogenetic analyses of <i>Acropora</i> Sox genes in ersion 2.3	28
Figure 2.2	Boxshade alignment of 79 amino acids HMG domain of SoxB genes 3	30
Figure 2.3	The schematic drawing of conserved regions of major Sox groups (B-F)	31
	Spatial expression patterns of AmSoxB1, AmSoxB3 and AmSoxE1 during yogenesis	
-	Spatial expression patterns of AmSoxC throughout the developmental	36
Figure 2.6	Cell morphology of AmSoxC expressing cells.	38
Figure 2.7	Spatial expression pattern of NvSoxC during Nematostella development4	10
Figure 3.1	The canonical Wnt/β-catenin signaling pathway	50
Figure 3.2	Spatial expression pattern of <i>Ambcat</i> mRNA	58
_	Inhibition of <i>Acropora</i> gastrulation by 5uM alsterpaullone (A.P) treatment	50
_	Gastrulation rates (%) of <i>Acropora</i> embryos treated with 5uM	60

Figure 3.5	Western blot a	nalysis of the GST- <i>Ambcat</i> fusion protein
Figure 3.6	Distribution of	β-catenin protein during <i>Acropora</i> early embryogenesis 63
Figure 3.7	The effect of a	lsterpaullone on the distribution of β-catenin protein 65
		rpaullone on the expressions of variety genes examined by
•		uM alsterpaullone on the spatial expression pattern of
-		5uM alsterpaullone on the spatial expression pattern of
-		SuM alsterpaullone treatment on the spatial expression
Supplementa	ry figure 2.1	Sequence analysis of AmSoxB1
Supplementa	ry figure 2.2	Sequence analysis of AmSoxB2
Supplementa	ry figure 2.3	Sequence analysis of AmSoxB3
Supplementa	ry figure 2.4	Sequence analysis of AmSoxC
Supplementa	ry figure 2.5	Sequence analysis of AmSoxE1
Supplementa	ry figure 2.6	Sequence analysis of AmSoxF
Supplementa genes examin	, ,	Temporal expression patterns of <i>Acropora millepora</i> Sox R
Supplementa	ry figure 2.8	Spatial expression pattern of AmSoxB1 after gastrulation107
* *	ry figure 2.9 al stages	Spatial expression pattern of AmSoxB2 throughout the
Supplementa	ry figure 2.10	Cell morphology of AmSoxB2 expressing cells 109
Supplementa	ry figure 2.11	Spatial expression pattern of AmSoxB3 after gastrulation109
Supplementa	ry figure 2.12	Spatial expression of AmSoxE1 after gastrulation 110
	ry figure 2.13	Spatial expression pattern of AmSoxF throughout
		Maximum Likelihood phylogenetic analyses of <i>Acropora</i> s in MolPhy version 2.3

Supplementary figure 2.15 Nematostella development (Martindale et al., 2004)113
Supplementary figure 2.16 Expression patterns for a subset of the <i>Nematostella</i> Sox genes (Magie et al., 2005)
Supplementary figure 2.17. Comparison of expression patterns between <i>Acropora</i> and <i>Nematostella</i> Sox genes during gastrulation
Supplementary figure 2.18. Comparison of the expression patterns between <i>Acropora</i> and <i>Nematostella</i> SoxC genes
Supplementary figure 3.1 No effect of alsterpaullone on the spatial expression pattern of <i>hbn-Am</i>
Supplementary figure 3.2 Boxshade alignment of the conserved short motif in C-terminal region of group E and F Sox genes

List of Tables

Table 1 Distribution of Sox gene groups in the Metazoa	8
Supplementary table 1 PCR primer set, sequences, PCR cycles and amplicon size for	
RT-PCR 11	7

Abstract

Members of the Sox transcription factor family have a wide variety of roles in the development of higher animals, including neural development and early embryogenesis. To better understand both the evolution of the Sox family and the roles of these genes in cnidarians, we are studying the Sox gene complement of the coral, Acropora millepora (Class Anthozoa). Based on overall domain structures and HMG box sequences, the Acropora Sox genes clearly fall into four of the five major Sox classes. Each of these genes has a clear Nematostella ortholog, but in most cases the expression pattern observed in Acropora differs significantly from that reported in Nematostella. AmSoxC is expressed in the ectoderm in a cell specific manner during development, with expression beginning much earlier than in Nematostella. During gastrulation, AmSoxB1 and AmSoxB3 transcripts are detected only in the presumptive ectoderm whereas AmSoxE1 transcription is restricted to the presumptive endoderm, suggesting that these Sox genes might play roles in germ layer specification. Again, the expression patterns reported for the corresponding Nematostella genes differ in many respects from those observed in Acropora. These differences may reflect diversity both in fundamental developmental processes and the underlying molecular mechanisms within the anthozoan Sub-Class Hexacorallia (Zoantharia).

Wnt/ β -catenin-signalling has important and multiple roles during early metazoan embryogenesis, including axial patterning or early embryogenesis. Upon receipt of the Wnt signal, β -catenin protein, which acts as a transcriptional regulator, is translocated into nuclei. Interactions between Sox proteins and β -catenin protein during germ layer formation have been reported, in a number of higher animals. To better understand the molecular mechanisms of early embryogenesis in *Acropora* and the ancestral roles of Wnt/ β -catenin signalling during early embryogenesis, the distribution of nuclear β -catenin protein was investigated during early development. In *Acropora*, whole-mount immunohistochemistry revealed that, unlike most other animals, including the sea anemone *Nematostella*, β -catenin protein accumulates in nuclei in the presumptive ectoderm of the blastula stage. At the 256–512-cell stage, β -catenin starts to accumulate in nuclei, and nuclear localization is observed in the presumptive ectoderm of the blastula stage. To investigate the function of β -catenin and its potential role as a

regulator of Sox gene expression during embryogenesis, Acropora embryos were treated with alsterpaullone, a specific inhibitor of the Wnt/ β -catenin signalling inhibitor GSK3 β . Alsterpaullone treatment significantly inhibited gastrulation in Acropora embryos and suppressed the expression of AmSoxB1 and AmSoxE1, suggesting that AmSoxB1 and E1 are downstream targets of Wnt/ β -catenin signalling. These results indicate that Wnt/ β -catenin signalling and several of the Sox genes play important roles in gastrulation and/or in germ layer formation in Acropora. As in the case of the Sox genes, the ectodermal accumulation of nuclear β -catenin in Acropora embryos again illustrates the surprisingly diversity of molecular mechanisms involved in early development of cnidarians.